Original Research Article

Evaluation of antibacterial activity of Commiphora myrrha against antibiotic resistant clinical pathogens

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ARTICLE INFO:

Article history:
Received: 25 June 2015
Received in revised form: 20 July 2015
Accepted: 18 August 2015
Available online: 30 September 2015

Keywords:
Antibacterial,
Commiphora myrrha,
MIC,
Oleogum resin

Abstract

There are many plants used by tribal people as an antibacterial, among them myrrh is the one which is commonly used. Myrrh is an oleo gum resin obtained from the plant Commiphora myrrha belongs to the family Burseraceae. It can adhere to intestines because of its resinous nature and it reduces the acidity in small intestines. Generally, the resin is collected from bark and stem of the plant by the process incision. In present study ethyl acetate extract of Commiphora myrrha was used for the evaluation of Anti bacterial activity against three Gram negative organisms and two Gram positive organisms. The method used in evaluation of Anti bacterial activity was serial dilution and Agar Diffusion method. The antibacterial activity was calculated in terms of minimum inhibitory concentration (MIC) and minimum bacterial concentration (MBC). The antibacterial activity was compared with marketed antibiotic.

Introduction

Myrrh is obtained from Commiphora myrrha (family: Burseraceae). Generally, the resin is collected from bark and stem of the plant by the process incision. Myrrh contains 9 to 17% volatile oil, 20 to 40% alcohol-soluble resin, and approximately 30 to 60% water-soluble gum. Some of the chief components in myrrh are sesquiterpenes. Sesquiterpenes are a large family of C15 isoprenoid molecules found in plants, microbes, and some marine organisms. Isoprenoids also called terpenoids are unsaturated hydrocarbons found in essential oils and oleorsins of plants. Mono terpenoids. Sesquiterpenoids present in volatile fraction and triterpenoids present in nonvolatile fraction. Along with these it also contains salt, sulphates, benzoic, malate and acetate, commiphoric acid, cinnamic aldehyde, furano eudesma 1, 3 diene. Main constituent attribute to its analgesic activity by binding to opioid receptors in brain. It is partly soluble in alcohol, water, and ether. It is soluble in excess of ethyl acetate and dimethyl sulphoxide. Diterpenoids having anti-bacterial activity

Antifungal and anti bacterial activity has been observed in vitro against standard pathogenic strains of Escherichia coli, Staphylococcus aureus, Pseudomonas aeruginosa and Candida albicans. The oleo-gum-resin of C. mukul was shown to be comparable to kanamycin against both Gram (+) and Gram (-) bacteria in vitro. Also proved to have carminative and anti-inflammatory. It is used internally to treat fevers, edema, anemia, constipation, hepatitis, ulcers, skin disorders, obesity, hemorrhoids, cough, asthma, paralysis, gout, and rheumatism. Anti-inflammatory activity, as observed in an animal model of inflammation. In this study, anti-inflammatory activity was more marked than that of hydrocortisone. An animal model of RA confirmed significant anti-inflammatory effects with oral administration, also resulting in decreased joint swelling. Several compounds found within myrrh exert local anaesthetic activity, chiefly by blocking the inward sodium current across membranes. Commiphora myrrha has many medicinal powers and has been used to treat various diseases, such as fever, ache, amenorrhea, tumors, stomach complaints, chest ailments, and skin infections in ancient India. Myrrh has astringent activity, promotes tissue granulation and enhances wound healing.

Materials and Methods

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Collection and authentication of plant sample
Commiphora myrrha was collected from local market of Hyderabad authenticated in Pharmacognosy lab of a Arya college of pharmacy. Voucher specimen is deposited in department; with voucher specimen number (ACP-G1). All the chemicals like ethanol ,dimethyl sulfoxide , sulphuric acid are of analytical grade.

The microorganisms were collected from the microbiology laboratory in Hyderabad viz. Gram positive organisms – Streptococcus aureus, Enterococci and Gram negative organisms - Escherichia coli , Pseudomonas aeruginosa, Klebsiella pneumoniae.

All experiments were carried out by standard published protocols. All glass wares and equipments used for handling of bacterial cultures were sterilized prior to use. All sterilization procedures were performed by autoclaving at 121°C at 15Lb pressure.

Methodology
Preparation of extract
The small pieces of oleogum resin of Commiphora myrrha was extracted three times with solvent of ethyl acetate at room temperature for 24hr with mass to volume ratio 1:10 g/ml and was extracted three times with solvent of ethyl acetate at room temperature for 24hr with mass to volume ratio 1:10 g/ml and was evaporated to dryness under vacuum at 40°C. All extracts were filtered to 0.45um membrane filters. For anti bacterial activity dried residue of extract was dissolved in DMSO (dimethyl sulfoxide)

Antibacterial activity of plant extract: The following different growth medias were used to culture the microorganisms.

Media for Bacterial Culture
Composition of Muller Hinton agar medium: peptone-5 grams/litre, beef extract-1.5 grams/litre, yeast-1.5 grams/litre, sodium chloride-5 grams/litre, agar agar-15 grams/litre, PH-7.3 at 25°C suspended 35 gm in 1000 ml of water. Composition of Nutrient broth: Enzymatic digested gelatine -8 gm, beef extract-5 gm, final PH 25°C suspended 13 gm in 1000 ml of water.

Preparation of Media: Accurately weighed the Muller Hinton agar medium, transferred in to clean conical flask and volume made up to 200ml with continuous shaking. Heated to dissolve the media in the distilled water. The conical flask was tightly closed with cotton plug and autoclaved at 121°C at 15Lb pressure.

Preparation of Nutrient Broath: Accurately weighed 13gm of nutrient broth and transferred in to clean conical flask and made the volume up to 1000 ml with continuous shaking. prepared broth was dispensed in to glass tubes each 10 ml and sealed tightly with cotton plugs. Tubes were autoclaved at 121°C for 20 minutes at 15Lb pressure.

Preparation of culture: All bacterial strains were obtained from microbial type culture collection. Different bacterial strains were maintained on nutrient agar and subcultures were freshly prepared before use.

Bacterial cultures were prepared by two methods
1. Plate Preparation: Using sterile conditions 20 ml of agar medium was transferred into sterile petri dishes. After solidifying, the plates were used for the assay.
2. Sub-Culturing: Subcultures were prepared by transferring loop full of inoculum from culture slants to freshly prepared agar slants. These were incubated in the desired conditions.

Determination of Antibacterial Activity
The Broth Dilution Method
Preparation of Microorganisms
The strains were maintained on sterilized Mueller-Hinton agar slants at 4°C. A loop full of each bacterial strain from agar slants was inoculated into 50 ml of sterile Nutrient broth in 100 ml conical flask which was sterilized at 121°C for 20 min at 15Lb pressure. The flasks were incubated at 37°C for 4-6hrs to activate the strain. Flasks containing the bacterial strain in broth were kept at 4°C for storage.

Determination of Minimum Inhibitory Concentration (MIC) and Minimum Bacterial Concentration (MBC)
MIC and MBC of extracts were determined by tube broth dilution assay. A sterile 2-fold dilution, ranging from 50mg - 0.39 mg/ml of extract prepared and 500 ul of each dilution was incubated with 2.47 ml of nutrient broth containing 30 µl of inoculum at 37°C for 24hrs. The MIC was determined at lowest concentration that demonstrates no visible growth by macroscopic evaluation. After determination of MIC, tubes showing no turbidity were diluted 100-fold with drug free nutrient broth and incubated at 37°C for 48 hrs. The lowest concentration of tube that showed no visible growth in drug free cultivation was considered as the MBC.

Determination of Antibacterial Activity by Agar Well Diffusion Method [6]
An agar-well diffusion method was employed for evaluation of antibacterial activity. The bacterial strains were reactivated from stock cultures by transferring into nutrient broth and incubating at 37°C for 18 hrs. A final inoculums containing 10^6 colony forming units (1x10^6 CFU/ml) was added to wells (8 mm in diameter) punched on agar surface. Plates were incubated overnight at 37°C and diameter of inhibition zone (DIZ) around each well was measured in mm. Antibiotics at the concentration of 100ug/well were used as positive reference to determine sensitivity of microorganisms tested.

Results and Discussion
Antibacterial activity by dilution method: The antibacterial activity of ethyl acetate extract of C. myrrha showed in Table-I. the MIC and MBC was calculated by observing growth and turbidity of nutrient broth.
Table 1: Determination of MIC and MBC of plant extract against different microorganisms (mg/ml)

<table>
<thead>
<tr>
<th>Organisms</th>
<th>MIC (mg/ml)</th>
<th>MBC (mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Escherichia coli</em></td>
<td>12.5</td>
<td>25</td>
</tr>
<tr>
<td><em>Enterococcus</em></td>
<td>3.12</td>
<td>12.5</td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td>6.25</td>
<td>12.5</td>
</tr>
<tr>
<td><em>Klebsiella pneumoniae</em></td>
<td>6.25</td>
<td>12.5</td>
</tr>
<tr>
<td><em>Streptococcus aureus</em></td>
<td>6.25</td>
<td>12.5</td>
</tr>
</tbody>
</table>

The results obtained in the present study showed that MIC against *Enterococci* is high i.e. 3.12mg/ml.

**Agar well diffusion method:** The antibacterial activity was done by using different concentrations of *C. myrrha* extract on different organisms shoed in table-2. Zone of inhibition was measured in mm.

Table 2: Agar well diffusion of plant extract against the different microorganisms (mg/ml)

<table>
<thead>
<tr>
<th>Organism</th>
<th>100 mg/ml</th>
<th>50 mg/ml</th>
<th>25 mg/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Escherichia coli</em></td>
<td>10 mm</td>
<td>6 mm</td>
<td>0</td>
</tr>
<tr>
<td><em>Enterococcus</em></td>
<td>17 mm</td>
<td>12 mm</td>
<td>10 mm</td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td>11 mm</td>
<td>10 mm</td>
<td>0</td>
</tr>
<tr>
<td><em>Klebsiella pneumoniae</em></td>
<td>10 mm</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td><em>Streptococcus aureus</em></td>
<td>9 mm</td>
<td>5 mm</td>
<td>0</td>
</tr>
</tbody>
</table>

Five bacterial strains were used to evaluate the effect of plant extract by agar well diffusion method. In all the organisms the highest zone of inhibition was observed for *Enterococci* at 100 mg/ml (17 mm). And no inhibition zone found for *Klebsiella pneumoniae* at 50mg and 25mg. Finally the plant extract shows the inhibition zones at 100mg, 50mg, 25mg, against the organism *Enterococci*. For remaining organisms less or no inhibition zones were found. So *Enterococci* organism is more sensitive to *Commiphora myrrha*.

The zone of inhibition obtained for different organisms is presented in Figure-1.

**Figures 1:** Agar well diffusion method of plant extract against the different microorganisms (mg/ml)
Comparison study of antibacterial activity of plant extract with antibiotic on clinical pathogens by Agar well diffusion method

The Comparison study of antibacterial activity of *C. myrrha* with marketed antibiotic was performed on different organisms and showed in table-3. The zone of inhibition obtained for different organisms is presented in figure-2.

**Table 3: Comparison study of antibacterial activity of plant extract with antibiotic on clinical pathogens**

<table>
<thead>
<tr>
<th>Organisms</th>
<th>Extract (200mg/ml)</th>
<th>Antibiotic (100mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Klebsiella pneumoniae</em></td>
<td>15 mm</td>
<td>26 mm</td>
</tr>
<tr>
<td><em>Streptococcus aureous</em></td>
<td>10 mm</td>
<td>25 mm</td>
</tr>
<tr>
<td><em>Enterococcus</em></td>
<td>22 mm</td>
<td>30 mm</td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>17 mm</td>
<td>24 mm</td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td>12 mm</td>
<td>40 mm</td>
</tr>
</tbody>
</table>

Agar well diffusion method of cefataxime sodium, a cephalosporin antibiotic shown the highest zone of inhibition against *Pseudomonas aeruginosa* at 100 mg/ml (40 mm) while as *Commiphora myrrha* extract shows the highest zone of inhibition against *Enterococci* at 200 mg/ml (22mm).

**Figures 2: Comparison of antibacterial activity of c.myrrha with marketed antibiotic on clinical pathogens by Agar well diffusion method**

1. *Streptococcus aureous*  
2. *Klebsiella pneumoniae*  
3. *E.coli*  
4. *Enterococcus*
Conclusion

In this study Commiphora myrrha was extracted and its biological activity was tested against antibacterial resistant organisms. The study shows that the organism Enterococci shows high sensitivity to Commiphora myrrha plant extract. The minimum inhibitory concentration (MIC) and MBC according to tube dilution method was found to be 3.12mg/ml and 12.5mg/ml respectively. Further the antibacterial activity of extract was compared with antibiotic like cefataxime against different organisms. The extract shows the high antibacterial activity against Enterococci organism where as cefataxime shows the high antibacterial activity against Pseudomonas aureginosa. The extract shows high zone of inhibition against Enterococci (17mm) by agar well diffusion method.

Many herbs are being used by the tribunal people. The scientific data should be produced to explore the activities of these herbal drug. In this research a small attempt has been made to explore the activity of C.myrrha, such kinds of study will surely help to explore the claims of herbal drugs.

Conflict of interest: We declare that we have no conflict of interest.

References
